

# Autoradiographic localization of $\beta$ -adrenoceptors in pig lung using [ $^{125}\text{I}$ ]-iodocyanopindolol

R.G. Goldie, J.M. Papadimitriou\*, J.W. Paterson, P.J. Rigby & D. Spina

Departments of Pharmacology and Pathology\*, University of Western Australia, Nedlands, Perth, 6009, Western Australia

1 The binding of the  $\beta$ -adrenoceptor radioligand [ $^{125}\text{I}$ ]-iodocyanopindolol (I-CYP) has been studied in pig lung parenchyma and the distribution of binding sites visualised by light microscopic autoradiography.

2 I-CYP binding was saturable (maximum binding capacity  $B_{\text{max}} = 51 \pm 3 \text{ fmol mg}^{-1} \text{ protein}$ ), involving sites with high affinity (dissociation constant  $K_D = 73 \pm 10 \text{ pM}$ ).

3 Specific I-CYP binding was displaceable both by  $\beta$ -adrenoceptor agonists ((–)-isoprenaline > (–)-adrenaline > ( $\pm$ )-fenoterol > (–)-noradrenaline > (+)-isoprenaline > ( $\pm$ )-RO363) and antagonists (( $\pm$ )-propranolol > ICI-118551 > atenolol), indicating a predominance of  $\beta_2$ -adrenoceptors. Further analysis showed that displacement data for the  $\beta_1$ -selective antagonist atenolol and the  $\beta_2$ -selective antagonist ICI-118551 were fitted best to a 2 binding site model and that both  $\beta_1$ - and  $\beta_2$ -adrenoceptors were present in pig lung in the ratio 28:72 respectively.

4 Autoradiographic grains were localized over tissue and were most dense over alveolar walls > vascular endothelium > vascular smooth muscle > bronchial smooth muscle = bronchial epithelium. Atenolol ( $10^{-5}\text{M}$ ) caused a 31% reduction in specific grain density over alveolar wall tissue, while a 10 fold lower concentration of ICI-118551 ( $10^{-6}\text{M}$ ) caused a 50% decrease. These results are consistent with binding data in pig lung parenchyma demonstrating a mixed population of  $\beta$ -adrenoceptors with a predominance of the  $\beta_2$  subtype.

5 Approximately 95% of the parenchymal  $\beta$ -adrenoceptors were associated with the alveolar wall as a mixed population of the  $\beta_1$  and  $\beta_2$  subtypes in the ratio 30:70 respectively. A greater proportion of the  $\beta$ -adrenoceptors associated with bronchial and vascular smooth muscle seemed to be of the  $\beta_2$  subtype.

6 It is possible that the previously described relaxant responses of the pig lung parenchyma strip to  $\beta$ -agonists, mediated via  $\beta_2$ -adrenoceptors, resulted from the sum of reactivities in airway and vascular smooth muscle together with relaxation of alveolar interstitial cells.

## Introduction

The lung parenchyma strip is a heterogeneous preparation containing bronchiolar, alveolar and vascular tissue, all of which may contribute to its contractility and relaxant responsiveness to drugs (Evans & Adler, 1981; Bertram *et al.*, 1983a). We have previously shown that  $\beta_2$ -adrenoceptors mediate relaxation of the pig lung parenchyma strip (Goldie *et al.*, 1982). However, both  $\beta_1$ - and  $\beta_2$ -adrenoceptors have been revealed in both membrane binding studies and in autoradiographic studies in lung parenchyma from the rat (Xue *et al.*, 1983), rabbit (Rugg *et al.*, 1978) and guinea-pig (Carswell & Nahorski, 1983). It was of interest to determine the distribution of  $\beta$ -adrenoceptors in pig lung parenchyma to identify those tissue structures upon which  $\beta_2$ -adrenoceptors

reside. It is these structures which must constitute that component of the pig lung strip which respond functionally to relaxant  $\beta$ -agonists. The possible existence of parenchymal  $\beta_1$ -adrenoceptors was also investigated.

## Methods

### Tissue preparation

Central lung lobes were obtained from pigs freshly slaughtered at a local abattoir. Lobes were inflated by tracheal instillation of OCT embedding medium diluted 1:4 with 0.9% w/v NaCl solution (saline).

Inflated lobes were snap frozen in isopentane, quenched with liquid nitrogen and tissue blocks (approximately  $10 \times 10 \times 10$  mm) cut and stored at  $-75^\circ\text{C}$  until required. Serial frozen tissue sections ( $10\ \mu\text{m}$ , autoradiographs;  $16\ \mu\text{m}$ , kinetic experiments) were cut at  $-30^\circ\text{C}$  from a total of 20 blocks from 7 separate lungs and mounted and thawed onto acid washed, gelatinised glass slides. Sections were stored at  $-75^\circ\text{C}$  for up to 2 weeks before use without loss of radioligand binding capacity.

#### Radioligand binding studies

Slides with tissue sections were incubated at  $22^\circ\text{C}$  for 5–180 min in Tris-HCl buffer (170 mM; pH 7.6) containing [ $^{125}\text{I}$ ]-iodocyanopindolol (I-CYP,  $\sim 2000\ \text{Ci mmol}^{-1}$ ,  $10\text{--}320\ \text{pM}$ ) and the protease inhibitor phenylmethylsulphonylfluoride (PMSF,  $10\ \mu\text{M}$ ). Specific binding of I-CYP to  $\beta$ -adrenoceptors was defined as I-CYP binding which was displaceable by  $200\ \mu\text{M}$  (–)-isoprenaline.

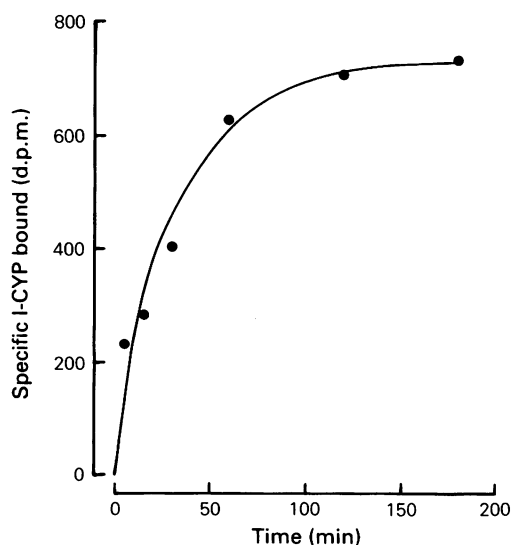
**Kinetic experiments** Lung sections ( $16\ \mu\text{m}$ ) were incubated with I-CYP ( $50\ \text{pM}$ ) for 150 min in the presence or absence of the  $\beta$ -adrenoceptor antagonists ( $\pm$ )-propranolol ( $10^{-9}\text{--}10^{-5}\ \text{M}$ ; non-selective), atenolol ( $6 \times 10^{-6}\text{--}10^{-2}\ \text{M}$ ;  $\beta_1$ -selective) or ICI-118551 ( $6 \times 10^{-8}\text{--}10^{-4}\ \text{M}$ ;  $\beta_2$ -selective) or the  $\beta$ -adrenoceptor agonists, (–)-isoprenaline ( $10^{-8}\text{--}10^{-4}\ \text{M}$ ), (+)-isoprenaline ( $10^{-6}\text{--}10^{-2}\ \text{M}$ ), (–)-noradrenaline ( $10^{-7}\text{--}10^{-3}\ \text{M}$ ), (–)-adrenaline ( $10^{-7}\text{--}10^{-4}\ \text{M}$ ), ( $\pm$ )-fenoterol ( $3 \times 10^{-8}\text{--}10^{-4}\ \text{M}$ ;  $\beta_2$ -selective) and RO363 ( $3 \times 10^{-5}\text{--}10^{-4}\ \text{M}$ ;  $\beta_1$ -selective). After incubation with ligands, sections were washed once in buffer at  $22^\circ\text{C}$  for 1 min and again for  $2 \times 15$  min periods and then quickly rinsed in distilled water. Sections were then wiped from slides with glass fibre filter paper (Whatman, GF/A) and the radioactivity counted in a Packard gamma counter (Model 5650). Twelve tissue sections from blocks from 3 different lungs were used for the estimation of each data point. Protein content in adjacent sections in each tissue block was estimated by the method of Lowry *et al.*, (1951). Preliminary binding constants were obtained in the saturation experiments using Scatchard and Hill analyses. Final binding parameter estimates and their errors were estimated by non-linear least squares regression analysis using the computer programme NONLIN (Metzler, 1969). In competition experiments, parameter estimates were obtained for 1 and 2 site binding models using the programme MLAB (N.I.H., U.S.A.).

**Autoradiography** Tissue sections ( $10\ \mu\text{m}$ ) were incubated with I-CYP ( $50\ \text{pM}$ ) for 150 min in the absence or presence of  $200\ \mu\text{M}$  (–)-isoprenaline or the  $\beta$ -adrenoceptor antagonists atenolol ( $10^{-7}\text{--}10^{-4}\ \text{M}$ ) or

ICI-118551 ( $10^{-8}\text{--}10^{-5}\ \text{M}$ ) and washed as described for kinetic experiments. Sections were then rapidly dried under a stream of cold dry air.

Coverslips (type 0) were coated at  $46^\circ\text{C}$  with NTB-3 nuclear track emulsion (Eastman Kodak Co., Rochester, N.Y.) diluted 1:1 with distilled water, air dried for 3 h and stored at  $4^\circ\text{C}$ . Emulsion coated coverslips were attached to one end of tissue slides with cyanoacrylate adhesive and stored in dessicated light-tight X-ray cassettes at  $4^\circ\text{C}$  for 3 days. The emulsion was developed in Dektol (Kodak) diluted 1:1 with distilled water for 3 min and fixed for 3 min with Rapidfix (Kodak) diluted 1:4 with distilled water. The tissue was stained with Gill's haematoxylin for 30 s. After air drying, the coverslips were re-apposed to the slides and the tissue mounted in DePeX medium (BDH). Sections were viewed with a Zeiss photomicroscope under light and dark-field illumination using a  $\times 10$  objective.

Quantitation of I-CYP binding sites in pig lung parenchyma was achieved by manual counting of autoradiographic grains visualised on dark-field photomicrographs of fields described over tissue areas of alveolar wall, airway and vascular smooth muscle, airway epithelium and vascular endothelium. Three hundred and forty-two fields, each with an area of approximately  $1000\ \mu\text{m}^2$  were chosen within 28 tissue sections from a pig lung. Sections were labelled with I-CYP in the absence or presence of  $200\ \mu\text{M}$  (–)-



**Figure 1** Time-dependent increase in specific [ $^{125}\text{I}$ ]-iodocyanopindolol (I-CYP;  $50\ \text{pM}$ ) binding in  $16\ \mu\text{m}$  sections of pig lung parenchyma. Each point represents the mean of data from 2 separate lung samples (4 tissue sections per lung sample).

isoprenaline, or the  $\beta$ -adrenoceptor antagonists ICI-118551 and atenolol.

The probability ( $P$ ) of differences between mean values for autoradiographic grain densities was determined using Student's two-tailed, non-paired  $t$  test and was considered significant if  $P < 0.05$ .

### Drugs

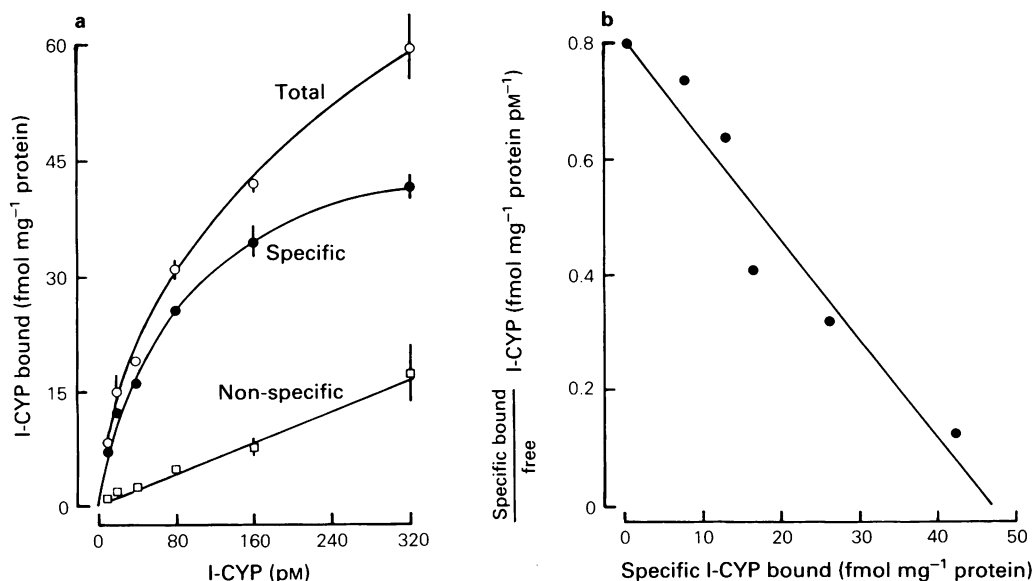
Drugs used were (–)-3-[ $^{125}$ I]-iodocyanopindolol (Amersham); (–)-isoprenaline hydrochloride, (+)-isoprenaline bitartrate, (–)-noradrenaline bitartrate, (–)-adrenaline bitartrate, (±)-propranolol hydrochloride (Sigma); RO363 ((±)-1-(3,4-dimethoxyphenylethylamino)-3-(3,4-dihydroxyphenoxy)-2-propanol)oxalate; synthesized at the Victorian College of Pharmacy, Melbourne, Australia); atenolol, ICI-118551 (erythro-DL-1-(7-methylindan-4-yloxy)-3-(isopropylaminobutan-2-ol) hydrochloride; ICI). Stock solutions of the catecholamines were freshly prepared in saline containing ascorbic acid  $20 \mu\text{g ml}^{-1}$ .

## Results

### Characteristics of I-CYP binding

The specific binding of I-CYP in pig lung parenchyma

reached equilibrium at between 120 and 180 min (Figure 1). Subsequent binding experiments were performed using an equilibration time of 150 min. Binding of I-CYP was saturable, involving a single population of non-interacting sites (mean  $\pm$  s.e. mean Hill coefficient,  $n_H = 0.933 \pm 0.052$ ). Specific binding sites had a high affinity for I-CYP (dissociation constant,  $K_D = 73.4 \pm 10.0 \text{ pM}$ ) and the maximum binding capacity ( $B_{\text{max}}$ ) was  $51 \pm 3 \text{ fmol mg}^{-1}$  protein (Figure 2). Each data point on the saturation curve is the mean of 12 observations derived from sections from 3 separate lung samples. Specific binding of I-CYP in pig lung parenchyma was displaceable by both  $\beta$ -agonists and antagonists (Figure 3). The potency order for displacement as determined by concentrations of competing  $\beta$ -agonists producing 50% reduction in specific binding ( $\text{IC}_{50}$ ) was (–)-isoprenaline > (–)-adrenaline > (±)-fenoterol > (–)-noradrenaline > (+)-isoprenaline > (±)-RO363. Data for (+)-fenoterol is not shown in Figure 3. The potency order for  $\beta$ -antagonists was (±)-propranolol > ICI-118551 > atenolol. Competition for specific I-CYP binding sites by isoprenaline was stereo-selective with the laevo isomer approximately 270 times more potent than the dextro isomer. (–)-Adrenaline was 4 times more potent than (–)-noradrenaline indicating the predominance of  $\beta_2$ -adrenoceptors. Displacement data for the  $\beta_1$ -selective antagonist atenolol and for the  $\beta_2$ -selective antagonist ICI-118551, were fitted best to

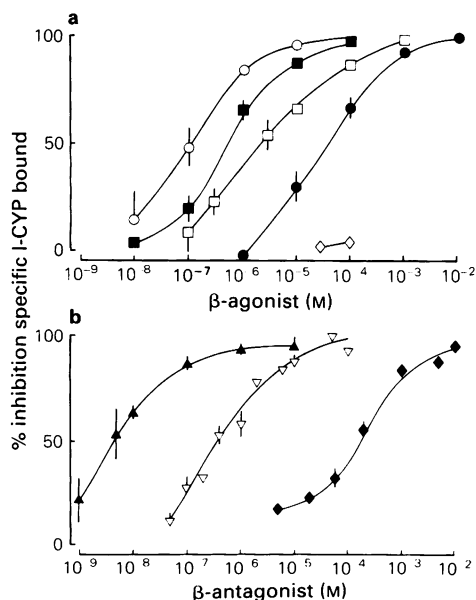


**Figure 2** (a) Saturation of [ $^{125}$ I]-iodocyanopindolol (I-CYP) binding in  $16 \mu\text{m}$  sections of pig lung parenchyma. Each point represents the mean of data from 3 separate lung samples (4 tissue sections per lung sample) and vertical lines indicate s.e. mean. Non-specific I-CYP binding was taken as that occurring in the presence of  $200 \mu\text{M}$  (–)-isoprenaline. Specific binding = total minus non-specific binding. (b) Scatchard analysis of specific I-CYP binding.

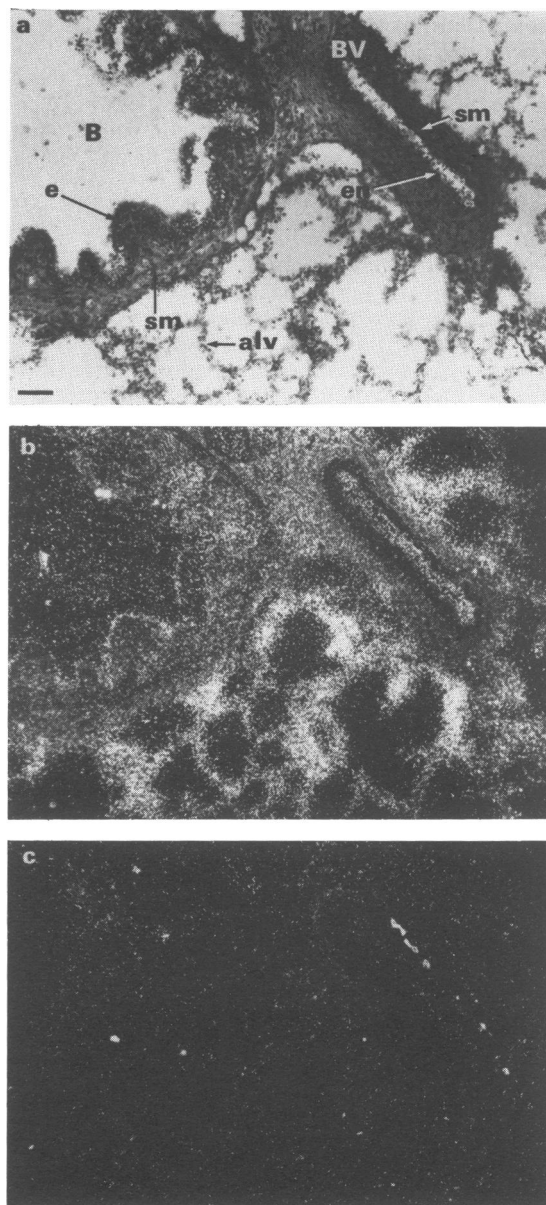
a 2 binding site model indicating that both  $\beta_1$ - and  $\beta_2$ -adrenoceptors were present in pig lung. Hill coefficients significantly less than unity were obtained for atenolol ( $n_H = 0.426 \pm 0.053$ ) and for ICI-118551 ( $n_H = 0.721 \pm 0.100$ ). Computer analysis (MLAB) of displacement data for atenolol showed that  $\beta_1$ - and  $\beta_2$ -adrenoceptors were present in the proportions  $31.4 \pm 8.1\%$  and  $68.6 \pm 8.2\%$  respectively. ICI-118551 displacement data determined the proportions as  $25.3 \pm 12.4\%$   $\beta_1$  and  $74.7 \pm 12.3\%$   $\beta_2$ . Thus on average, 28% of the  $\beta$ -adrenoceptors were of the  $\beta_1$  subtype, while 72% were  $\beta_2$ -adrenoceptors.

### Autoradiography

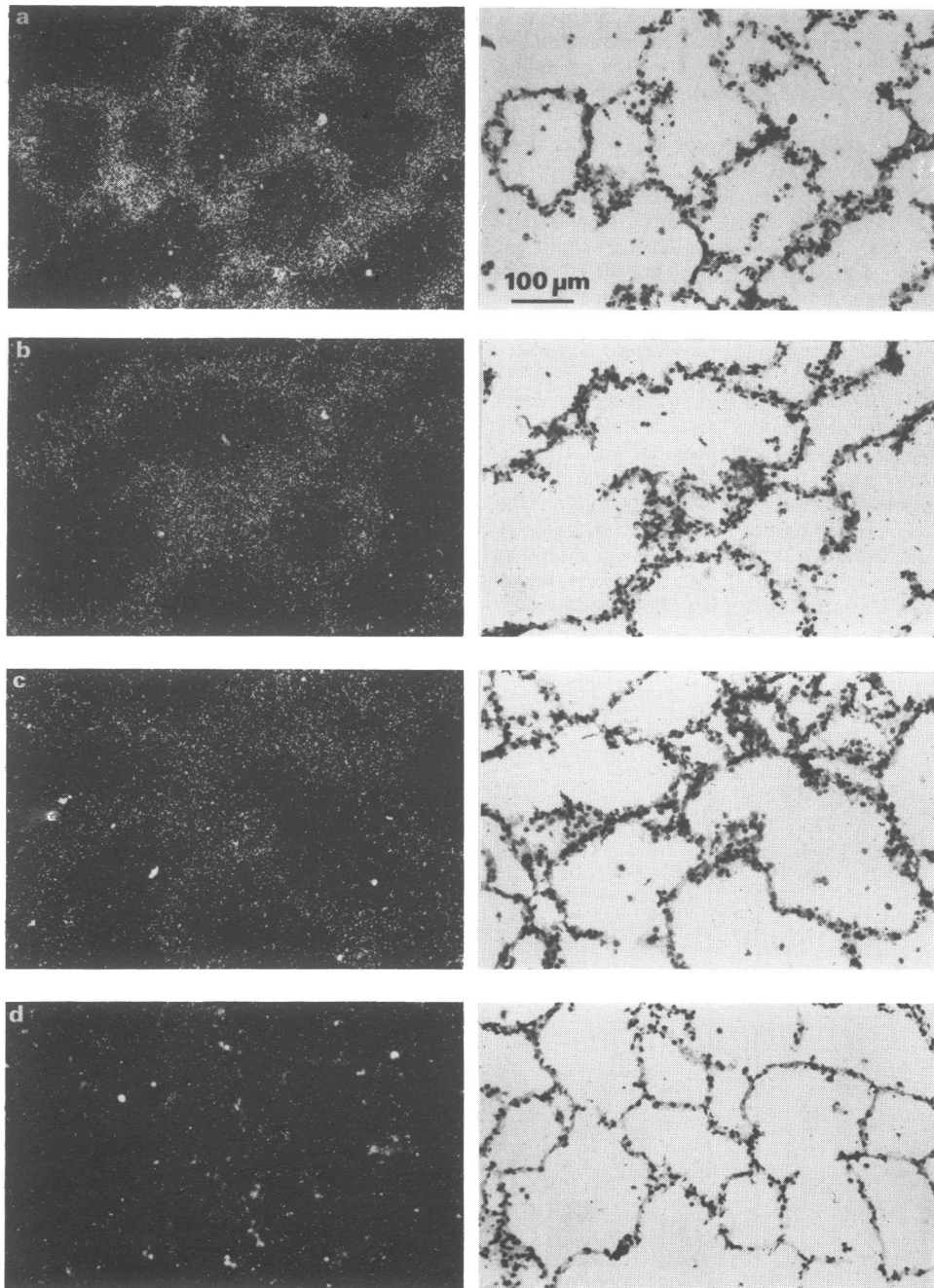
Autoradiographic grains representing I-CYP binding sites, were localized with different densities over several tissue types within the lung parenchyma including alveolar wall, vascular and airway smooth muscle, vascular endothelium and airway epithelium (Figure 4). Labelling over alveolar walls was dense and



**Figure 3** Concentration-effect relationships for the displacement of specific [ $^{125}$ I]-iodocyanopindolol (I-CYP; 50 pM) binding in 16  $\mu$ m sections of pig lung parenchyma by: (a) the  $\beta$ -adrenoceptor agonists (+)-isoprenaline (●), (-)-isoprenaline (○), (-)-noradrenaline (□), (-)-adrenaline (■) and (±)-RO363 (◇); (b) the  $\beta$ -adrenoceptor antagonists (±)-propranolol (▲), ICI-118551 (▽) and atenolol (◆). Each point represents the mean of data from at least 3 separate lung samples (4 tissue sections per lung sample).



**Figure 4** (a) Bright-field photomicrograph of a 10  $\mu$ m frozen section of pig lung parenchyma. B = bronchiole, BV = blood vessel, e = epithelium, en = endothelium, sm = smooth muscle, alv = alveolar wall. Bar = 100  $\mu$ m. (b) Dark-field photomicrograph of the above section showing the distribution and localization of autoradiographic grains derived from [ $^{125}$ I]-iodocyanopindolol (I-CYP, 50 pM) binding. (c) Dark-field photomicrograph showing the distribution of non-specific autoradiographic grains in the next serial section incubated with I-CYP (50 pM) and (-)-isoprenaline (200  $\mu$ M).

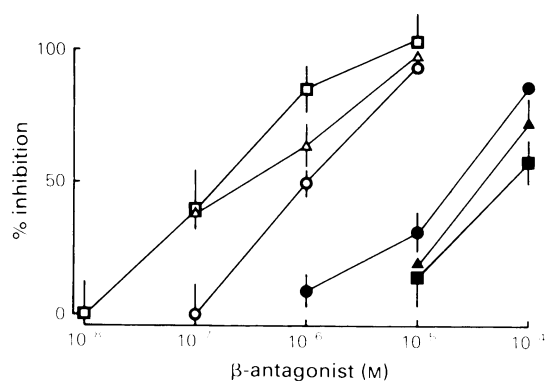


**Figure 5** Dark-field (left-hand panels) and bright-field (right-hand panels) photomicrographs of 10  $\mu$ m frozen sections of pig lung parenchyma showing the localization of autoradiographic grains derived from [ $^{125}$ I]-iodocyanopindolol (I-CYP, 50 pM) binding in alveolar wall in the absence (a), or the presence of (b) atenolol,  $10^{-5}$ M; (c) ICI-118551,  $10^{-6}$ M or (d) (-)-isoprenaline, 200  $\mu$ M. Bar = 100  $\mu$ m.

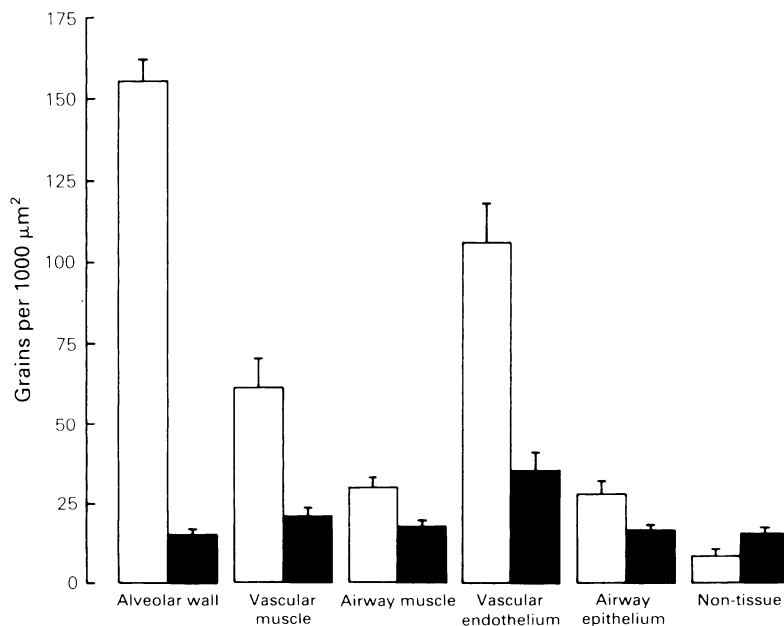
uniform, giving no indication that I-CYP binding was selective for particular cell types. Sections incubated with I-CYP in the presence of 200  $\mu\text{M}$  (–)-isoprenaline demonstrated low levels of uniform non-specific binding.

The effects of the  $\beta$ -adrenoceptor antagonists atenolol ( $\beta_1$ -selective) and ICI-118551 ( $\beta_2$ -selective) on total I-CYP binding in alveolar wall tissue are shown in Figure 5. Atenolol ( $10^{-5}\text{M}$ , Figure 5b) and ICI-118551 ( $10^{-6}\text{M}$ , Figure 5c) caused reductions in specific grain density of  $30.8 \pm 6.7\%$  and  $49.9 \pm 4.3\%$  respectively. Atenolol and ICI-118551 reduced grain counts over tissue to background levels at  $10^{-3}\text{M}$  and  $10^{-5}\text{M}$  respectively, concentrations where non-selective displacement was expected.

The relative densities of  $\beta$ -adrenoceptors (as determined by autoradiographic grain counting) were estimated over alveolar wall, vascular endothelium and smooth muscle, and airway epithelium and smooth muscle (Figure 6). Alveolar wall tissue was the most densely labelled tissue type followed by vascular endothelium, although this was 32% lower than that over alveolar wall. The grain density over vascular smooth muscle was 61% lower than that over alveolar wall tissue, but was twice as high as that over airway



**Figure 7** Concentration-effect relationships between the  $\beta_1$ -selective antagonist atenolol (solid symbols), the  $\beta_2$ -selective antagonist ICI-118551 (open symbols) and reduction in the densities of autoradiographic grains (% inhibition) derived from [ $^{125}\text{I}$ ]-iodocyanopindolol (I-CYP, 50 pM) binding in alveolar wall tissue (●,○), airway smooth muscle (■,□) and vascular smooth muscle (▲,△) in a sample of pig lung. Vertical lines represent s.e.mean of 5–14 field measurements.



**Figure 6** Mean densities of specific (open columns) and non-specific (filled columns) autoradiographic grains (grains per 1000  $\mu\text{m}^2$ ) derived from [ $^{125}\text{I}$ ]-iodocyanopindolol (I-CYP, 50 pM) binding in alveolar wall tissue (specific,  $n = 44$  fields; non-specific,  $n = 28$  fields), vascular ( $n = 14;9$ ) and airway smooth muscle ( $n = 24;18$ ), vascular endothelium ( $n = 15;9$ ), airway epithelium ( $n = 24;16$ ) and over non-tissue (airway lumen;  $n = 24;37$ ) in a sample of pig lung. Vertical bars on columns represent s.e.mean.

smooth muscle and epithelium. Grain densities over connective tissue elements, including bronchial cartilage, were very low and approximately equivalent to those over non-tissue areas.

The effects of atenolol ( $10^{-7}$ – $10^{-4}$ M) and of ICI-118551 ( $10^{-8}$ – $10^{-5}$ M) on I-CYP binding were assessed in terms of reductions in autoradiographic grain densities over alveolar wall tissue and airway and vascular smooth muscle (Figure 7). ICI-118551 was 10 times and 5 times more potent in airway and vascular smooth muscle respectively, than in alveolar wall tissue. Conversely, atenolol was 2–3 times less potent in these tissues than in alveolar wall. These data are consistent with the existence of both  $\beta_1$ - and  $\beta_2$ -adrenoceptors in alveolar wall tissue, while more homogeneous populations of  $\beta_2$ -adrenoceptors exist in airway and vascular smooth muscle.

## Discussion

The present study documents the tissue localization of  $\beta$ -adrenoceptors in pig lung parenchyma using the potent, highly selective radioligand I-CYP (Engel *et al.*, 1981; Munzel *et al.*, 1983). Kinetic experiments showed that I-CYP was bound to a saturable population of sites with high affinity. Both  $\beta$ -adrenoceptor-selective agonists and antagonists displaced specific I-CYP binding. Furthermore, competition for I-CYP binding sites by isoprenaline was stereoselective, in that the laevo isomer was 270 times more potent than the dextro isomer, a finding similar to that of Carstairs *et al.*, (1984) in human lung. Taken together, these results indicate that specific I-CYP binding in pig lung was selective for  $\beta$ -adrenoceptors.

The heterogeneous nature of lung tissue was reflected in the different autoradiographic grain densities observed over different tissue types within the lung. The highest grain density was associated with alveolar wall tissue, as previously observed in both human (Carstairs *et al.*, 1984) and rat (Conner & Reid, 1984) lung. In contrast, in the ferret (Barnes *et al.*, 1982) and rabbit (Barnes *et al.*, 1984), bronchial smooth muscle contained a significantly greater density of  $\beta$ -adrenoceptors than did alveolar wall tissue. The density of  $\beta$ -adrenoceptors over vascular smooth muscle was twice that observed over airway smooth muscle and epithelium. While epithelial  $\beta$ -adrenoceptors do not appear to be directly relevant to the expression of mechanical responses of airway smooth muscle they are known to mediate increases in the active transport of chloride ions (Davis *et al.*, 1979), increases in the velocity of airway mucous transport (Mossberg *et al.*, 1976a,b) and to influence epithelial cell differentiation (Jones & Reid, 1979). Very low levels of specific binding were detected over connective tissue elements and cartilage. With the exception of the low grain density measured

over airway epithelium, the relative grain densities over particular tissue types in pig lung were similar to those determined in human lung (Carstairs *et al.*, 1984).

Competition binding experiments demonstrated that  $\beta_2$ -adrenoceptors predominate in pig lung parenchyma. However, further analysis of data from the present study, describing the displacement of specific I-CYP binding by atenolol and ICI-118551 revealed that  $\beta_1$ -adrenoceptors constituted 28% of the total population. Manual counting of autoradiographic grains indicated that the density of  $\beta$ -adrenoceptors was approximately 2.5 times and 5 times greater over alveolar wall tissue than over vascular and airway smooth muscle respectively. Volume density analysis has previously shown that human lung parenchyma is 78% alveolar wall, 8% vascular smooth muscle and only 3% airway smooth muscle, with the remaining 11% consisting mainly of cartilage and connective tissue elements (Bertram *et al.*, 1983a,b).

Assuming that the composition of pig lung parenchyma is similar to human lung parenchyma, approximately 95% of the total population of  $\beta$ -adrenoceptors must have been associated with the alveolar wall. This is in close agreement with the value of 97% established in rat lung alveolar wall (Conner & Reid, 1984). Given that approximately 28% of the total population of parenchymal  $\beta$ -adrenoceptors were of the  $\beta_1$  subtype, it is clear that the great majority of these were associated with the alveolar wall. Thus alveolar tissue contained both  $\beta_1$ - and  $\beta_2$ -adrenoceptors approximately in the proportion 30:70, respectively. Autoradiographic grain counting demonstrated a smaller potency difference between ICI-118551 and atenolol in alveolar tissue than in bronchial and vascular smooth muscle (Figure 6). This indicates a higher percentage of  $\beta_2$ -adrenoceptors in smooth muscle than in alveolar tissue.

We have previously shown that  $\beta$ -agonist-induced relaxation of the pig lung parenchyma strip was mediated via  $\beta_2$ -adrenoceptors (Goldie *et al.*, 1982). It is often assumed that relaxation of lung strips reflects the reactivity of airway smooth muscle rather than of vascular or alveolar components. However, in view of the very low volume density of airway smooth muscle in lung parenchyma as well as the relatively sparse population of  $\beta_2$ -adrenoceptors in the peripheral airways, it seems likely that relaxant response of peripheral blood vessels may also contribute to  $\beta$ -agonist-induced relaxation of the lung parenchyma strip.

Our data suggest that the majority of alveolar  $\beta$ -adrenoceptors are located on cells which do not subserve mechanical responses. This is consistent with data showing that  $\beta$ -agonists stimulate the release of surfactant from alveolar type II cells (Dobbs & Mason, 1979; Giannopoulos, 1980) and may regulate fluid absorption from the alveolar spaces in the new-

born (Walters & Olver, 1978). However, a small population of potentially contractile alveolar interstitial cells has been described in several species of lung (Kapanci *et al.*, 1974) and these may contain  $\beta_2$ -adrenoceptors. Since the majority of the lung parenchyma tissue consists of alveolar wall, such interstitial cells may also contribute to the relaxant responsiveness of the lung strip to  $\beta$ -agonists.

## References

- BARNES, P.J., BASBAUM, C.B., NADEL, J.A. & ROBERTS, J.M. (1982). Localization of beta-adrenoceptors in mammalian lung by light microscopic autoradiography. *Nature*, **299**, 444–447.
- BARNES, P.J., JACOBS, M. & ROBERTS, J.M. (1984). Glucocorticoids preferentially increase fetal alveolar beta-adrenoceptors: Autoradiographic evidence. *Ped. Res.*, **18**, 1191–1194.
- BERTRAM, J.F., GOLDIE, R.G., PAPADIMITRIOU, J.M. & PATERSON, J.W. (1983a). Correlations between pharmacological responses and structure of human lung parenchyma strips. *Br. J. Pharmac.*, **80**, 107–114.
- BERTRAM, J.F., GOLDIE, R.G., PAPADIMITRIOU, J.M. & PATERSON, J.W. (1983b). Responses of peripheral human lung to serotonin and norepinephrine and their relationships to bronchiolar and vascular volume densities. *Acta. Stereol.*, **2**, (Suppl. 1), 261–264.
- CARSTAIRS, J.R., NIMMO, A.J. & BARNES, P.J. (1984). Autoradiographic localisation of beta-adrenoceptors in human lung. *Eur. J. Pharmac.*, **103**, 189–190.
- CARSWELL, H. & NAHORSKI, S.R. (1983). Autoradiographic localisation of  $\beta$ -adrenoceptors in guinea-pig airways. *Br. J. Pharmac. Proc. Suppl.*, **80**, 50P.
- CONNER, M.W. & REID, L.M. (1984). Mapping of beta-adrenergic receptors in rat lung. Effect of isoproterenol. *Exp. Lung Res.*, **6**, 91–101.
- DAVIS, B., MARIN, M.G., YEE, J.W. & NADEL, J.A. (1979). Effect of terbutaline on movement of  $\text{Cl}^-$  and  $\text{Na}^+$  across the trachea of the dog *in vitro*. *Am. Rev. Resp. Dis.*, **120**, 547–552.
- DOBBS, L.G. & MASON, R.J. (1979). Pulmonary alveolar type II cells isolated from rats: Release of phosphatidylcholine in response to beta-adrenergic stimulation. *J. clin. Invest.*, **63**, 378–387.
- ENGEL, G., HOYER, D., BERTHOLD, R. & WAGNER, H. (1981).  $(\pm)$ - $[^{125}\text{I}]$ -cyanopindolol, a new ligand for beta-adrenoceptors: Identification and quantitation of subclasses of beta-adrenoceptors in guinea-pig. *Naunyn-Schmiedeberg's Arch. Pharmac.*, **317**, 277–285.
- EVANS, J.N. & ADLER, K.B. (1981). The lung strip: Evaluation of a method to study contractility of pulmonary parenchyma. *Exp. Lung Res.*, **2**, 187–195.
- GIANNOPOULOS, G. (1980). Identification and ontogeny of beta-adrenergic receptors in fetal rabbit lung. *Biochem. biophys. Res. Commun.*, **95**, 388–394.
- GOLDIE, R.G., PATERSON, J.W. & WALE, J.L. (1982). A comparative study of  $\beta$ -adrenoceptors in human and porcine lung parenchyma strip. *Br. J. Pharmac.*, **76**, 523–526.
- JONES, R. & REID, L.M. (1979). Beta-agonists and secretory cell number and intracellular glycoprotein in airway epithelium, the effect of isoproterenol and salbutamol. *Am. J. Pathol.*, **95**, 407–422.
- KAPINCI, Y., ASSIMACOPOULOS, A., IRLE, C., ZWAHLEN, A. & GABBIANI, G. (1974). Contractile interstitial cells in pulmonary alveolar septa. A possible regulator of ventilation/perfusion ratio? *J. Cell Biol.*, **60**, 375–392.
- LOWRY, O.H., ROSEBROUGH, N.J., FARR, A.L. & RANDALL, R.J. (1951). Protein measurement with the folin phenol reagent. *J. biol. Chem.*, **193**, 265–275.
- METZLER, C.M. (1969). *A users manual for NONLIN and associated programs*. Kalamazoo, Michigan, U.S.A: The Upjohn Company.
- MOSSBERG, B., STRANDBERG, K., PHILLIPSON, K. & CAMNER, P. (1976a). Tracheobronchial clearance in bronchial asthma: Response to beta-adrenoceptor stimulation. *Scand. J. Resp. Dis.*, **57**, 119–128.
- MOSSBERG, B., STRANDBERG, K., PHILLIPSON, K. & CAMNER, P. (1976b). Tracheobronchial clearance and beta-adrenoceptor stimulation in patients with chronic bronchitis. *Scand. J. Resp. Dis.*, **57**, 281–289.
- MUNZEL, P.A., HEALY, D.P. & INSEL, P.A. (1983). Autoradiographic localisation of beta-adrenergic receptors in rat kidney slices using  $[^{125}\text{I}]$ -iodocyanopindolol. *Am. J. Physiol.*, **246**, F240–F245.
- RUGG, E.L., BARNETT, D.B. & NAHORSKI, S.R. (1978). Coexistence of  $\beta_1$  and  $\beta_2$  adrenoceptors in mammalian lung: Evidence from direct binding studies. *Mol. Pharmac.*, **14**, 996–1005.
- WALTERS, D.V. & OLVER, R.E. (1978). The role of catecholamines in lung liquid absorption at birth. *Ped. Res.*, **12**, 239–242.
- XUE, Q.-F., MAURER, R. & ENGEL, G. (1983). Selective distribution of beta and  $\alpha_1$ -adrenoceptors in rat lung visualized by autoradiography. *Arch. int. Pharmacodyn. Ther.*, **266**, 308–314.

(Received November 20, 1985.  
Accepted February 11, 1986.)